

Application Note

Using Isocratic Preparative Liquid Chromatography as a Quick First-Step Purification for Natural Product Isolation

Jo-Ann M. Jablonski, Rui Chen

Waters Corporation



Abstract

In this application note, a workflow to systematically develop isocratic prep LC methods for target compound isolation from complex natural product mixtures is demonstrated. The workflow starts with a generic LC analysis from which the elution %B of the target compound is calculated. The target compound is then isolated from the sample using an isocratic preparative LC method run at the calculated %B, leading to the collection of target compound with purities ranging from 73% to 100%. The isocratic prep LC methods were also proven rugged and reproducible. Compared to the commonly used gradient methods, the isocratic prep chromatography dramatically reduced the total run time and solvent usage, resulting in improved overall productivity and cost-effectiveness of the purification process. The proposed workflow is ideally suited as a quick, first step in a multi-step process for target compound isolation from natural products.

Benefits

- Isocratic preparative chromatography is ideally suited as a quick cleanup step prior to more stringent purification steps for complex mixtures, such as natural products.
- While attaining a target compound of reasonable purity, isocratic preparative chromatography also leads to shorter run time and reduced solvent usage, thereby improving throughput and the cost-effectiveness of the purification process.
- Isocratic preparative chromatography is rugged and reproducible.

Introduction

Preparative chromatography is the technique of choice for isolation and purification of different types of compounds, including pharmaceutical drug candidates, target compounds from synthetic mixtures, and bioactive compounds from natural products. As a result, there has been continuous research interest in improving the fundamental knowledge and instrumentation of preparative chromatography, with an ultimate goal of obtaining pure target compounds efficiently and cost-effectively.¹⁻⁶

Most purification strategies start with chromatographic profiling of the crude mixture at the analytical scale using either gradient HPLC or UPLC, followed by a scale-up to the semi-prep or preparative level. The most straightforward method transfer from analytical to prep chromatography is a direct geometric scale-up of a

linear gradient. Alternatively, a focused gradient approach, where a shallow gradient encompasses the elution %B of the target compound, is used for target compound isolation. While generally applicable, these two approaches often lead to long cycle time and increased solvent usage. Furthermore, depending on the sample complexity and desired purity, an iterative process might still be required to attain the target compounds. This is particularly true for natural product isolation, where a multi-step purification procedure is generally required due to the complexity of the sample.

This application note describes the use of isocratic chromatography as a first purification step to quickly and efficiently simplify the mixture and enrich the target compound prior to more stringent purification steps. Eriocitrin,⁷ a polar compound found in peppermint extract (Figure 1), was used as a model to demonstrate the isolation and purification workflow. The purity and solvent usage for each of the different purification approaches were compared. To ensure the general applicability of the proposed workflow, the methodology was further validated by isolating carnosic acid⁸ (Figure 1), a relatively non-polar compound from rosemary extract.

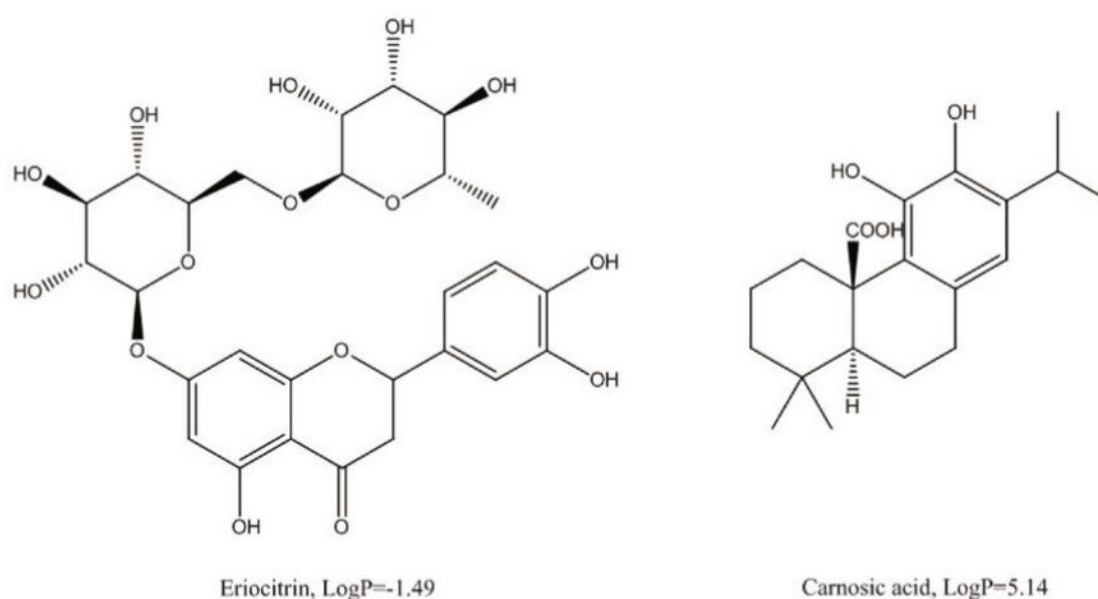


Figure 1. Chemical structures and LogP values of the two compounds used in this study including eriocitrin (MW = 596.5), and carnosic acid (MW = 332.4).

Experimental

Sample description

Eriocitrin:	A total of 4.9 g dried peppermint was extracted with 40 mL of 80:20 methanol/water solution for 72 hours at room temperature, with stirring. The supernatant was filtered with an Acrodisc Syringe Filter with GHP Membrane, 25 mm, 0.45 μm . The final extract volume was 29 mL.
Columns:	XSelect CSH C ₁₈ 5 μm , 4.6 x 100 mm XSelect C ₁₈ , OBD Prep 5 μm , 19 x 100 mm
Mobile phase A:	0.1% trifluoroacetic acid in water
Mobile phase B:	0.1% trifluoroacetic acid in acetonitrile
Detection:	220 nm
Flow rate:	Analytical, 1.46 mL/min; Prep, 25 mL/min
Gradient:	See Table 1
Injection volume:	See figures

Analytical		Prep									
		Direct scaling		Focused gradient		Isocratic without column washing		Isocratic with column washing		Isocratic loading at 0% B	
Time (min)	%B	Time (min)	%B	Time (min)	%B	Time (min)	%B	Time (min)	%B	Time (min)	%B
0.0	5.0	0.0	5.0	0.0	5.0	0.0	23.0	0.0	23.0	0.0	0.0
12.0	95.0	0.3	5.0	0.3	5.0	5.0	23.0	5.0	23.0	1.0	0.0
14.0	95.0	12.3	95.0	1.3	17.4			5.5	95.0	1.2	23.0
14.2	5.0	14.3	95.0	12.0	25.4			11.2	95.0	6.2	23.0
20.0	5.0	14.5	5.0	12.5	95.0			11.7	23.0	6.8	95.0
		20.3	5.0	17.5	95.0			17.4	23.0	11.8	95.0
				17.7	5.0					12.3	0.0
				25.7	5.0					17.3	0.0

Table 1. LC methods used for the analysis and purification of eriocitrin from peppermint extract. The analytical flow rate was 1.46 mL/min, and the preparative flow rate was 25 mL/min.

Carnosic acid:

The extraction of carnosic acid was performed using supercritical fluid extraction (SFE) on a Waters MV-10 ASFE System controlled by ChromScope Software. A total of 1.9 g of ground rosemary was loaded into a 5-mL extraction vessel. The extraction was performed for 90 minutes with 6 mL/min CO₂ and 0.2 mL/min methanol at 80 °C and 300 bar extraction pressure. The effluent was collected into a 20-mL scintillation vial with a makeup flow of 0.25 mL/min of isopropanol. The 5.5 mL of cloudy yellow extract was filtered through an Acrodisc Syringe Filter with GHP Membrane, 13 mm, 0.45 µm. The final volume of the filtered extract was brought to 6.5 mL by adding 1 mL methanol.

Columns:

XBridge C₁₈ 5 µm, 4.6 x 150 mm

XBridge C₁₈ OBD Prep 5 μm, 19 x 150 mm

Mobile phase A: 0.1% formic acid in water

Mobile phase B: 0.1% formic acid in acetonitrile

Detection: 220 nm

Flow rate: Analytical, 1.46 mL/min; Prep, 25 mL/min

Gradient: See Table 2

Injection volume: See figures

Analytical		Isocratic prep	
Time	%B	Time	%B
0.0	5.0	0.0	70.0
18.0	95.0	7.0	70.0
21.0	95.0		
21.3	5.0		
30.0	5.0		

Table 2. LC methods used for the analysis and isolation of carnosic acid from rosemary.

The analytical flow rate was 1.46 mL/min, and the preparative flow rate was 25.00 mL/min.

Instrumentation

The SFE experiment was performed on a Waters MV-10 ASFE System controlled by ChromScope Software. All LC experiments were performed on a Waters AutoPurification System controlled by MassLynx Software.

Results and Discussion

Comparison of different preparative methods for eriocitrin isolation

Eriocitrin, a flavonone compound rich in peppermint, has shown therapeutic effects against inflammation, allergies, and hypertension.⁷ Figure 2 shows the HPLC/UV chromatogram of eriocitrin using a generic gradient. Due to its polarity (LogP=-1.49), the target compound eluted from the column at 3.67 minutes.

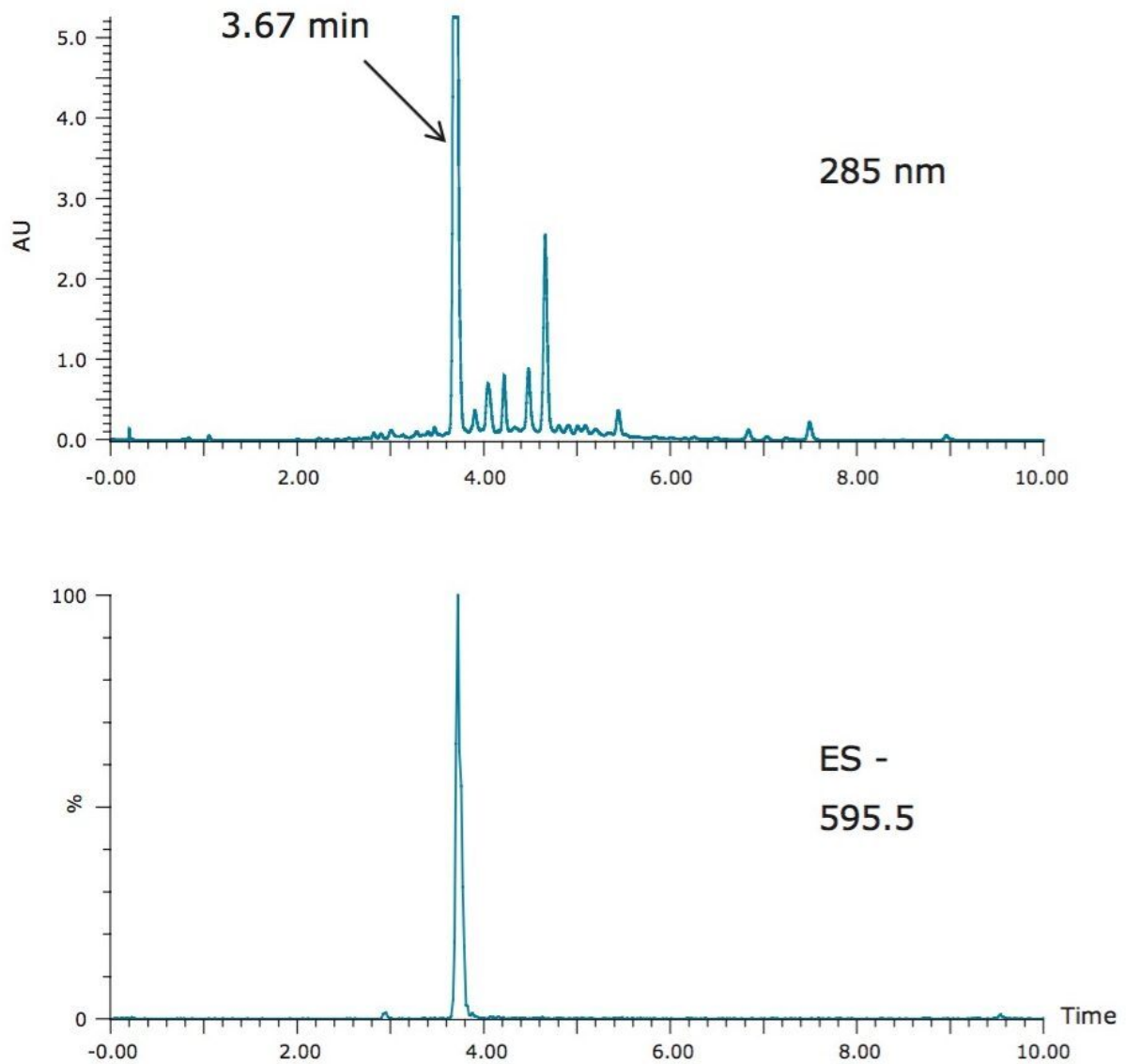


Figure 2. HPLC/UV chromatogram of the crude peppermint extract (top trace), and the extracted ion chromatogram (XIC) of eriocitrin (bottom trace). The injection volume was 10 μ L.

The most common approach for target compound isolation is direct geometric scaling of the analytical gradient to the preparative column. A representative prep chromatogram using the direct geometrical scale-

up is shown in Figure 3A. As noted in Table 1, the total run time for the preparative run was only slightly longer than the analytical one, due to the difference in system volume (primarily from the sample loop size difference) between the analytical and preparative systems.

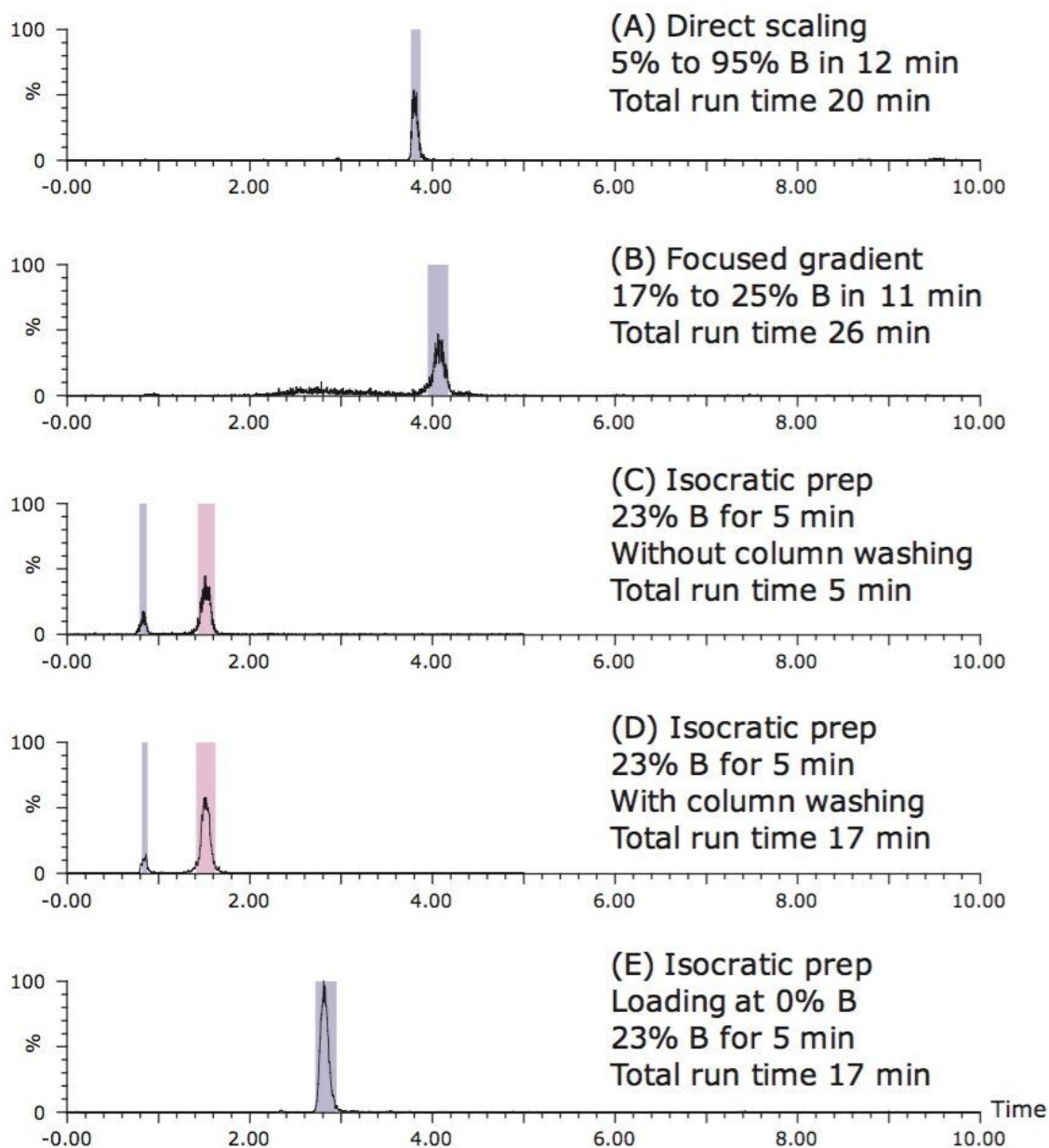


Figure 3. Comparison of the five preparative LC XICs of eriocitrin. All injection volumes were 171 μ L.

Alternatively, a focused-gradient approach⁹ can be used to isolate the target compound. Focused gradients are created with a shallow gradient segment centered around the %B where the target compound elutes from the column. A method for calculating the target compound elution percentage is as follows:

1. The system volume for the analytical LC system was experimentally determined to be 0.55 mL.

2. The column volume was determined to be 1.39 mL.

3. The volume between the point of gradient formation and the detector was, therefore,

$$0.55 \text{ mL} + 1.39 \text{ mL} = 1.94 \text{ mL}$$

4. The time required for the solvent to reach the detector: Time to the detector = offset in mL/Flow rate in mL/min = 1.94 mL/1.46 mL/min = 1.33 min

5. The actual time when the mobile phase composition responsible for peak elution was formed: Time actual composition formed = peak retention time – time to detector – gradient hold
= 3.67 min – 1.33 min – 0 = 2.34 min

6. The corresponding %B when the peak eluted:

$$\begin{aligned} \%B &= \text{Time actual composition formed} / \text{length grad segment} \times \text{change in gradient segment} + \text{initial gradient \%B} \\ &= 2.34 \text{ min} / 12 \text{ min} \times 90\% + 5\% \cong 23\% \end{aligned}$$

Based on the calculation, a focused gradient ranging from 17% B to 25% B in 10.7 minutes was developed. The slope of the focused gradient was 0.7% B per column volume, one-tenth of the original slope of 7.2% B per column volume. The shallow slope improved the resolution between the target compound and its adjacent peaks in the extract. In the preparative run using the focused gradient (Figure 3B), the target compound eluted off the column at ~4 minutes. The total run time for this method was 26 minutes, including column washing and equilibration.

The actual %B (23%) when the peak eluted in the analytical run was used for the isocratic preparative methods. Figures 3C through 3E show three different isocratic runs at 23% B. In addition to the eriocitrin peak at ~1.5 minutes, there was a small peak with a retention time of 0.8 minute, as shown in Figures 3C and 3D. This small peak contained a small amount of eriocitrin or its isomer, as well as other compounds. The presence of the small peak likely resulted from the breakthrough of the less retentive compounds, due to the loading at 23% B. To address the target compound loss due to the early eluting peak, the method was modified so that the initial loading condition was 0% B. After an initial one-minute hold at the loading condition, the %B was rapidly increased to 23% B. The resulting chromatogram is shown in Figure 3E. Clearly, the early eluting peak was eliminated. Injecting the sample at a lower %B in the method effectively mitigated the breakthrough of those less retentive compounds.

Table 3 summarizes the comparison of five different preparative methods for the eriocitrin purification. In

terms of purity, both gradient methods resulted in >95% purity of the target compound, while the three isocratic methods led to a similar purity of 89%. Incorporating a column washing and an equilibration step to the isocratic prep method increased the total run time to 17 minutes (Figures 3D and 3E), but did not lead to a significant improvement in target compound purity. With respect to solvent usage, all isocratic methods consumed less solvent than the gradient methods. Compared to the directly scaled gradient and focused gradient methods, the five-minute isocratic method reduced the solvent use by 88 and 78 mL/run, respectively.

Method	%B	Time (min)	% Purity (UV)	Calculated solvent B usage (mL/run)
Direct scale-up gradient	5 to 95	20	96	204
Focused gradient	17 to 25	26	100	194
Isocratic (without column wash)	23	5	89	116
Isocratic (with column wash)	23	17	89	177
Isocratic loading at 0% B (with column wash)	23	17	88	162

Table 3. Comparison of cycle time, purity, and calculated solvent B usage for different LC prep methods used for eriocitrin. *Average of three runs.

To gauge the ruggedness of the isocratic method, three replicates of the five-minute isocratic runs were performed. Figure 4 shows an overlay of three consecutive runs without column washing between injections. The results indicated that the isocratic methods were rugged and reproducible. Depending upon the assumed usage of the collected product, the described isocratic methods can be used as a high-throughput, first-step clean-up prior to more stringent purification, or as a final purification step. The isocratic methods improve productivity and cost-effectiveness due to the shortened run time and reduced solvent usage.

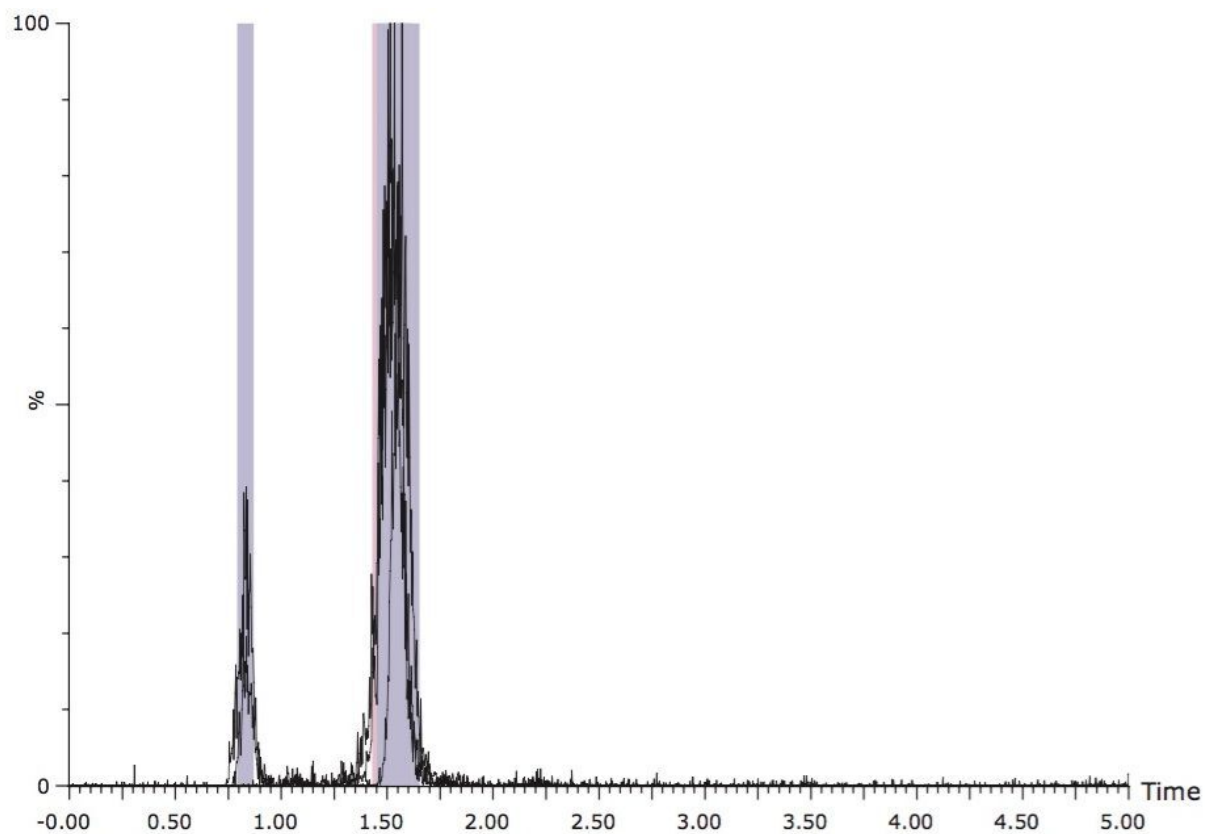


Figure 4. Overlaid XICs of three isocratic prep runs. The experimental conditions were the same as those in Figure 3C. The injection volumes were 171 $\mu\text{L}/\text{run}$.

Using a fast isocratic method for carnosic acid isolation

To ensure the general applicability of the described isocratic methodology, the same workflow was used for the isolation of carnosic acid, an anti-oxidant compound with a LogP of 5.14, found in the perennial herb, rosemary. Figure 5 shows the LC chromatograms of the rosemary SFE extract on an XBridge C₁₈ 5 μm , 4.6 x 150 mm Column, with a column volume of 2.093 mL. Note that a 30-minute screening gradient was used due to the column length (Table 2). The carnosic acid eluted at 14.9 minutes (Figure 5). Using the same procedure described in the previous section, the acetonitrile percentage at the elution of carnosic acid was determined to be 70%.

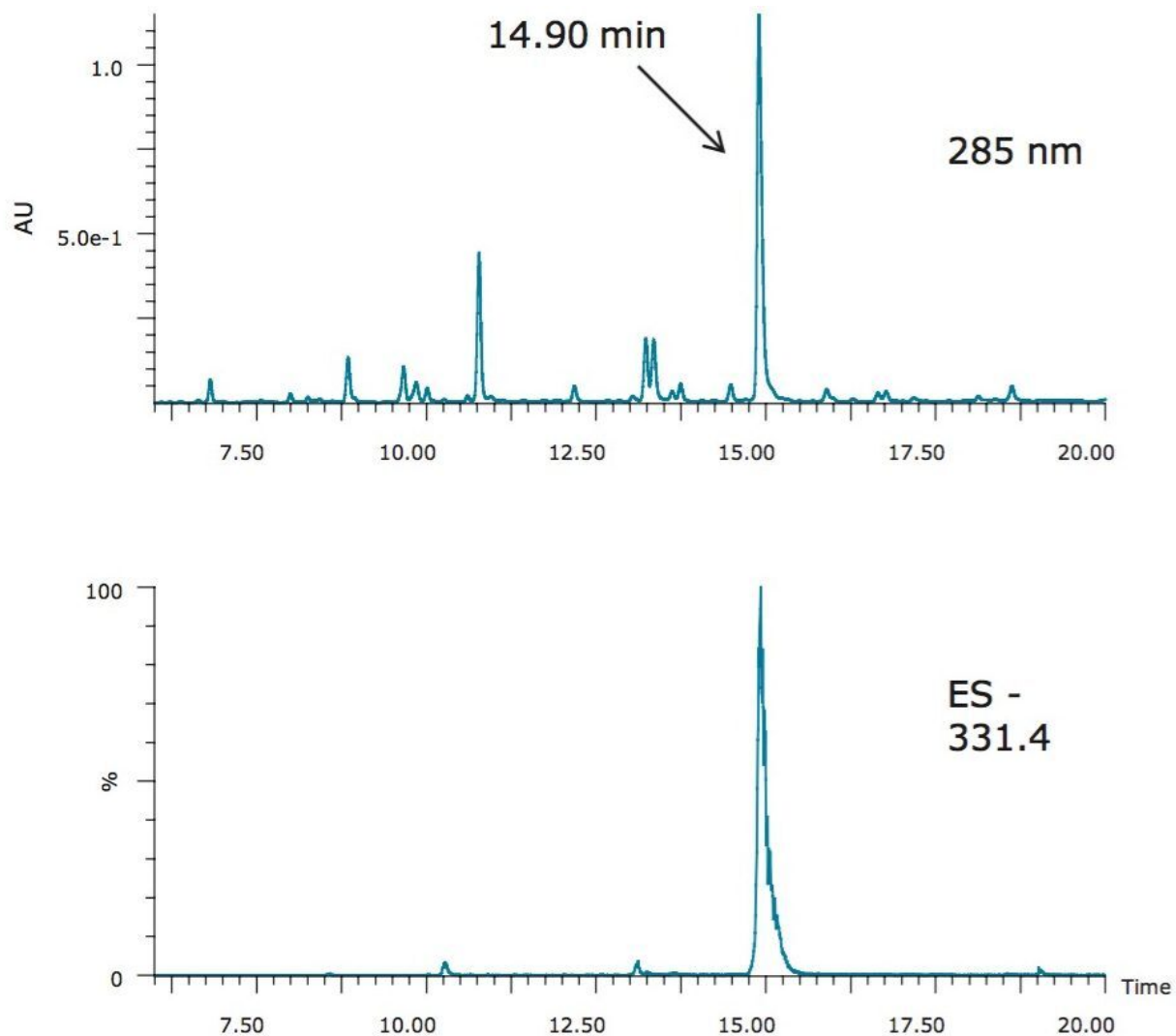


Figure 5. HPLC/UV chromatogram of the crude rosemary SFE extract (top trace), and the XIC of carnosic acid (bottom trace). The injection volume was 10 μ L.

Figure 6 shows the isocratic preparative chromatography of the rosemary SFE extract on an XBridge C₁₈ OBD Prep 5 μ m, 19 x 150 mm Column run at 70% B. Because of the volume of the fraction, the target compound was collected into two tubes. Fraction analysis (Figure 7) revealed that fraction 30 was 73% pure, and fraction 31 was 100% pure by UV. For fraction 30, the impurity peaks at 13.6 minutes and 14.5 minutes account for 2% and 25% of the total area, respectively. However, these two impurity peaks were well resolved from the carnosic acid. A second purification step would remove these impurities with relative ease.

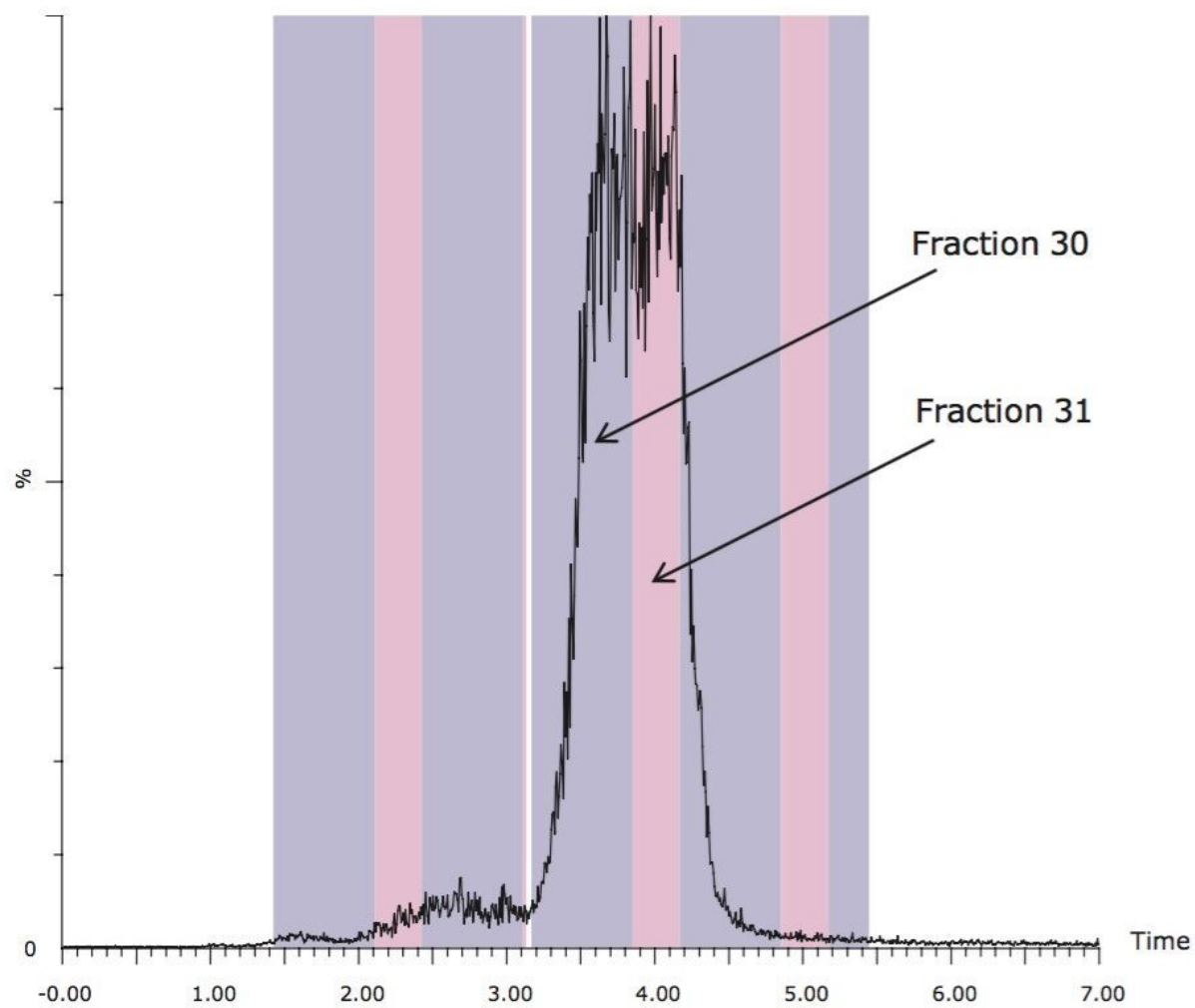


Figure 6. XICs of carnosic acid using isocratic prep chromatography. The injection volume was 1700 μ L.

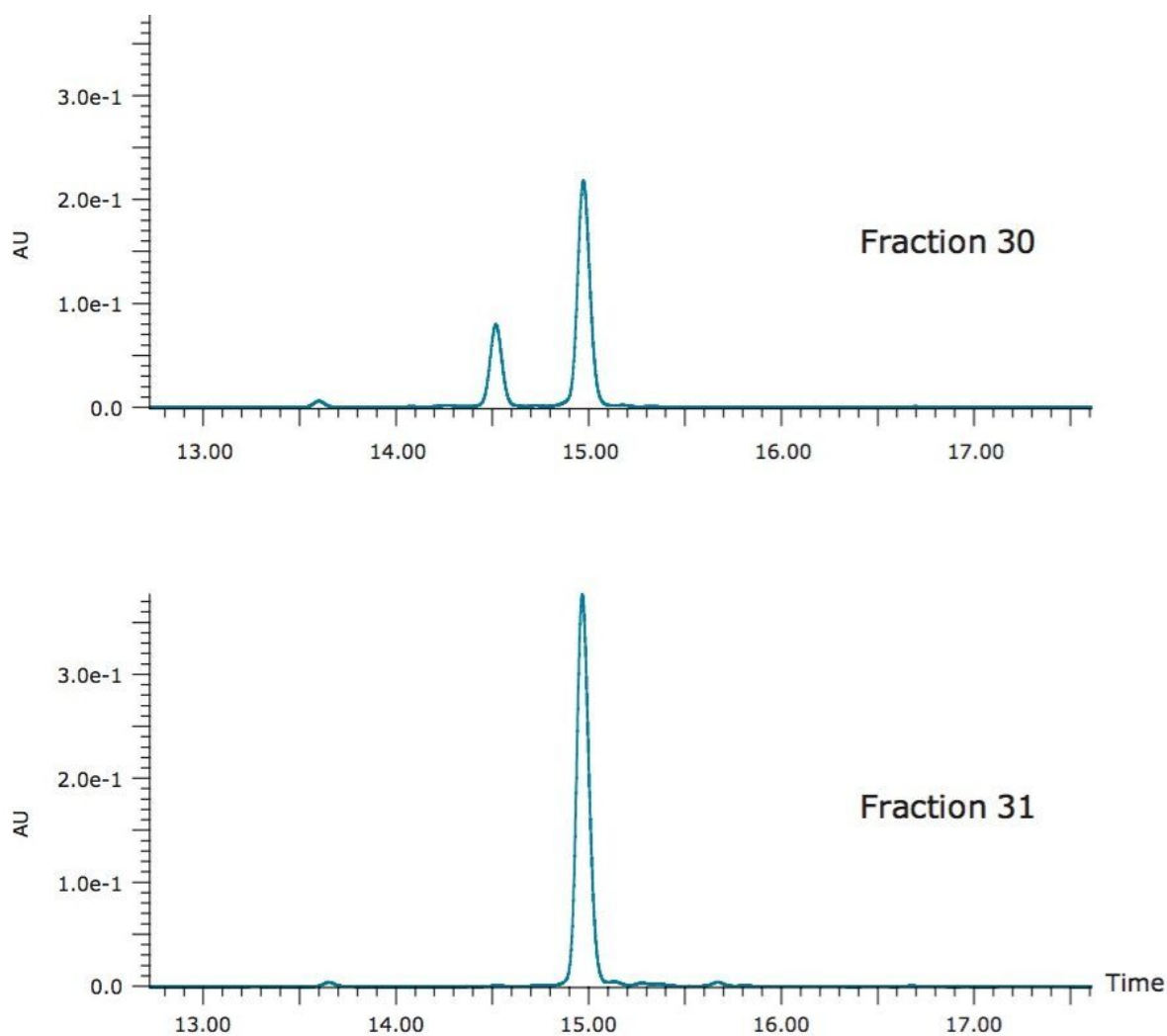


Figure 7. Analysis of two main fractions from the isocratic prep isolation of carnosic acid. The chromatographic conditions are listed in Table 2.

Conclusion

In this application note, a workflow to systematically develop isocratic prep LC methods for target compound isolation from complex natural product mixtures is demonstrated. The workflow starts with a generic LC analysis from which the elution %B of the target compound is calculated. The target compound is then isolated from the sample using an isocratic preparative LC method run at the calculated %B, leading to the collection of target compound with purities ranging from 73% to 100%. The isocratic prep LC methods were

also proven rugged and reproducible. Compared to the commonly used gradient methods, the isocratic prep chromatography dramatically reduced the total run time and solvent usage, resulting in improved overall productivity and cost-effectiveness of the purification process. The proposed workflow is ideally suited as a quick, first step in a multi-step process for target compound isolation from natural products.

References

1. Shan Y, Seidel-Morgenster A. Analysis of the Isolation of a Target Component using Multicomponent Isocratic Preparative Elution Chromatography. *Journal of Chromatography A*. 2004; 1041:53-62.
2. Guan H, Broyles BS, Guiochon G. Modeling of the Influence of the Experimental Conditions on the Separation of the Components of a Binary Mixture in Isocratic Overloaded Elution Preparative Chromatography. *Journal of Chromatography A*. 1996; 734:49-62.
3. Zhou T, Zhao W, Fan G, Chai Y, Wu Y. Isolation and Purification of Iridoid Glycosides from *Gardenia Jasminoides* Ellis by Isocratic Reversed-Phase Two-Dimensional Preparative High-Performance Liquid Chromatography with Column Switch Technology. *Journal of Chromatography B*. 2007; 858:296-301.
4. Wong V, Shalliker RA. Isolation of the Active Constituents in Natural Materials by 'Heart-Cutting' Isocratic Reversed-Phase Two-Dimensional Liquid Chromatography. *Journal of Chromatography A*. 2004; 1036:15-24.
5. Felinger A, Guiochon G. Comparing the Optimum Performance of the Different Modes of Preparative Liquid Chromatography. *Journal of Chromatography A*. 1998; 796:59-74.
6. Koza P, Gonnot V, Pelleter J. Right-First-Time Preparative Liquid Chromatography-Mass Spectrometry Purification. [dx.doi.org/10.1021/co200197b](https://doi.org/10.1021/co200197b) | *ACS Comb. Sci.* 2012; 14:273-279.
7. Fecka I, Turek S. Determination of Water-Soluble Polyphenolic Compounds in Commercial Herbal Teas from Lamiaceae: Peppermint, Melissa, and Sage. *J. Agric. Food Chem.* 2007; 55:10908-10917.
8. Herrero M, Plaza M, Cifuentes A, Ibanez E. Green Processes for the Extraction of Bioactives from Rosemary: Chemical and Functional Characterization via UltraPerformance Liquid Chromatography-Tandem Mass Spectrometry and In-vitro Assays. *Journal of Chromatography A*. 2010; 1217:2512-2520.
9. Jablonski JM, Wheat TE, Diehl DM. Developing Focused Gradients for Isolation and Purification. *Waters*

Featured Products

[AutoPurification System <https://www.waters.com/10007147>](https://www.waters.com/10007147)

[MV-10 ASFE System <https://www.waters.com/134661549>](https://www.waters.com/134661549)

[MassLynx MS Software <https://www.waters.com/513662>](https://www.waters.com/513662)

[ChromScope Software <https://www.waters.com/134647658>](https://www.waters.com/134647658)

720004693, May 2013